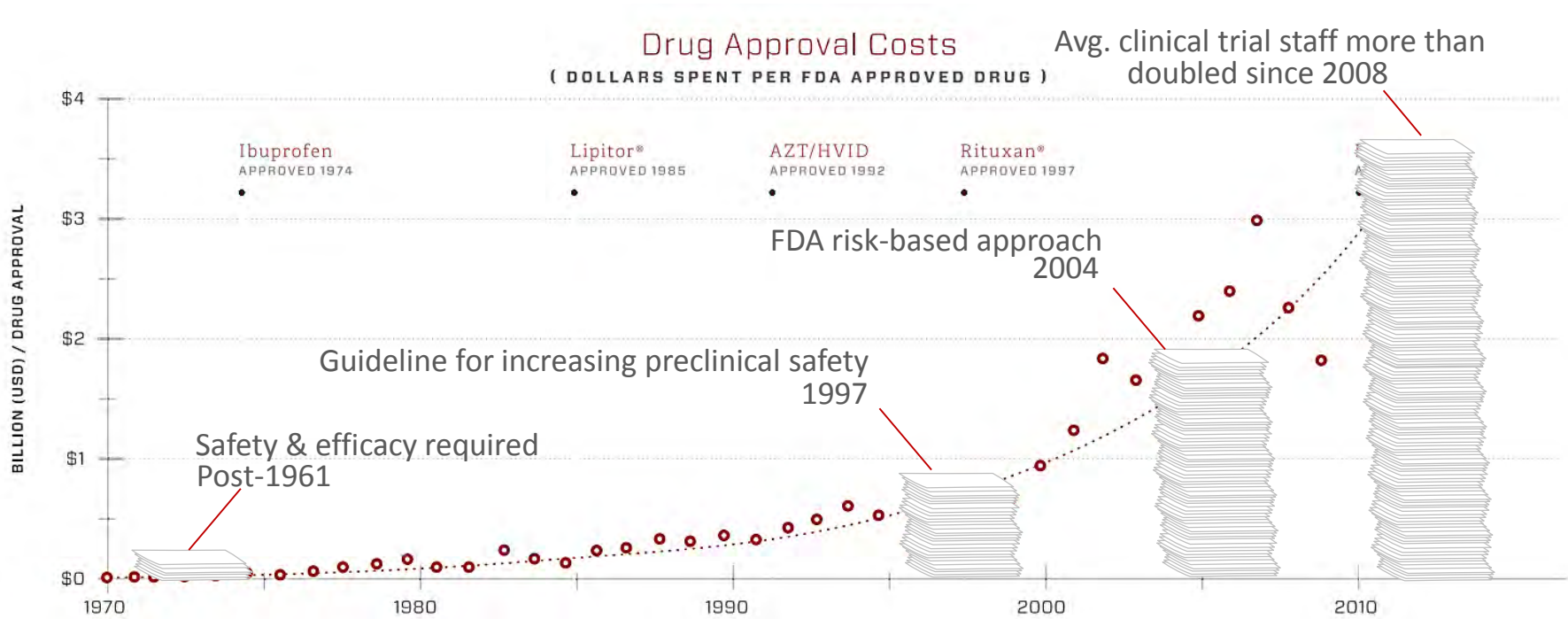


# **Human-on-a-Chip: a paradigm shift in substance testing!?**

# A “medieval” approach to drug development



Source: Cutting Edge Information *Clinical Trial Staffing Levels Skyrocket (2011)*  
Founders Fund *What Happened to the Future?*

# The Pharma Innovation Gap

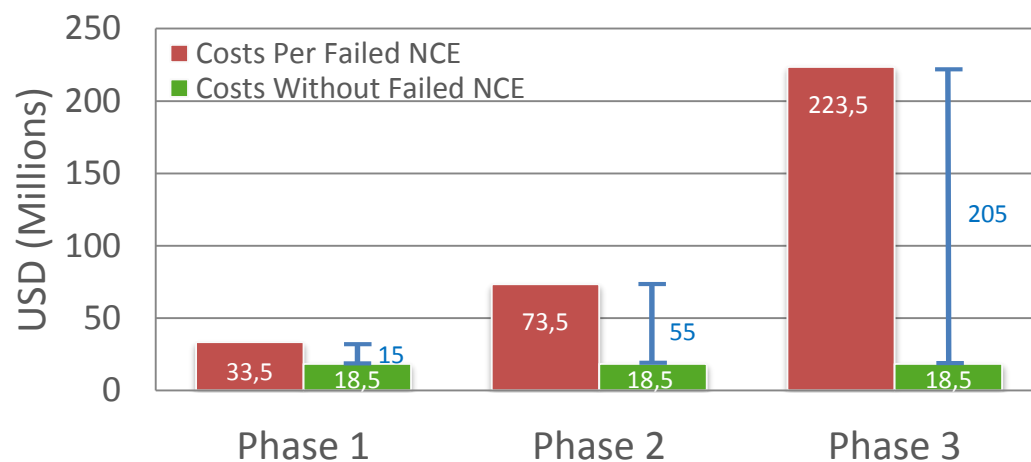
**Problem:** Drugs pass preclinical, but fail in clinical development

## Innovation Need:

*Of 100 New Chemical Entities (NCEs) progressed from preclinical to clinical development :*

- 46 fail due to toxicity
- 35 fail due to lack of efficacy

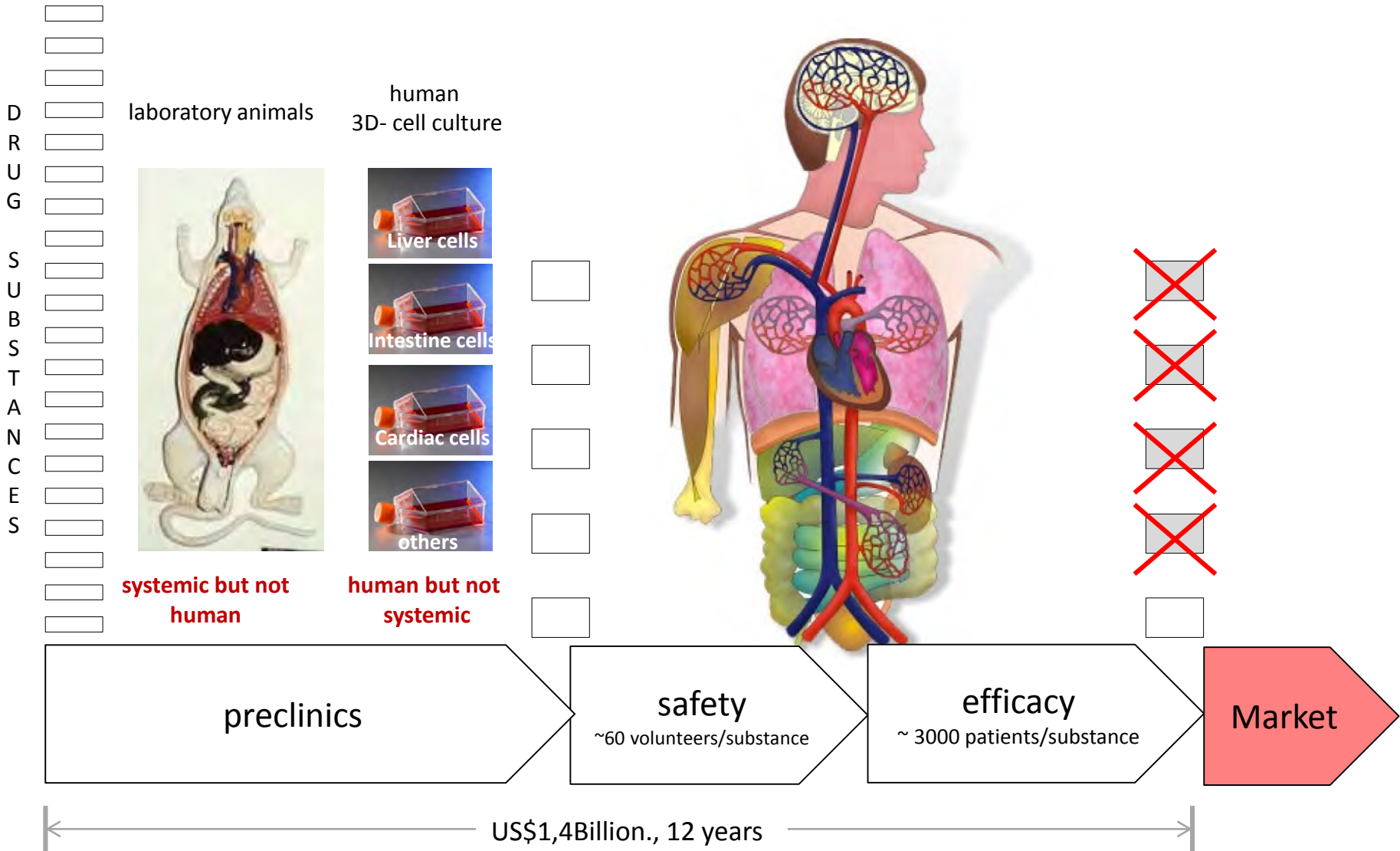
### Cost-Savings of Eliminating Failed NCEs



S. M. Paul et Al. How to improve R&D productivity: the pharmaceutical industry's grand challenge.

*Nature Reviews Drug Discovery*. 2010. (9), 205-214.

# Predictive power of preclinical testing



# Microfluidic-based homeostasis *in vitro*

number of  
“organs”

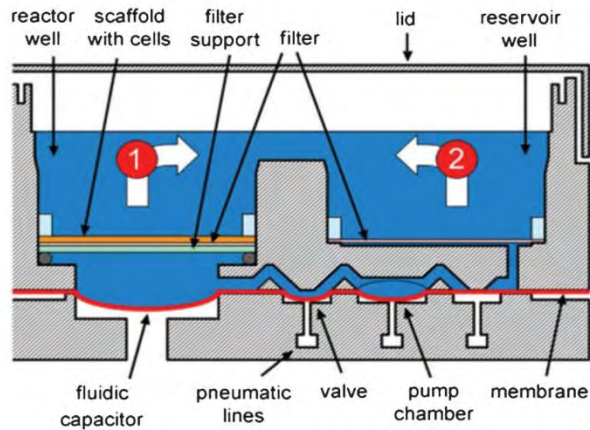
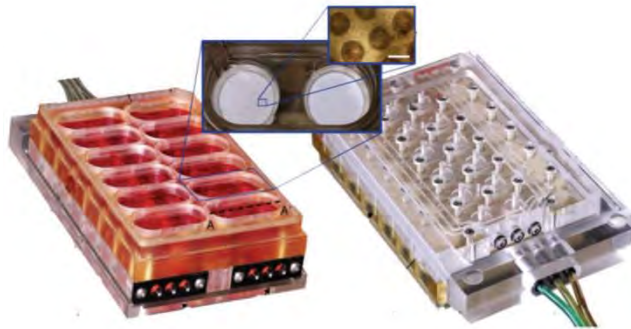
## microscale culture systems

	<u>homotopic</u>	<u>heterotopic</u>
<b>single</b>	<p><b>endothelial cells:</b> Young et al. 2010</p> <p><b>myoblasts:</b> Gu et al. 2004</p> <p><b>hepatocytes:</b> Powers et al. 2002, Leclerc et al. 2004, Ho et al. 2006, Lee et al. 2007, Toh et al. 2007 and 2009, Carraro et al. 2008, Park et al. 2008, Goral et al. 2010</p> <p><b>neurons:</b> Rhee et al. 2005</p> <p><b>mammary epithelial cells:</b> Grafton et al 2011</p> <p><b>adipose cells:</b> Nakayama et al. 2008</p> <p><b>embryo cells:</b> Hung et al. 2005, Chung et al. 2005, Villa-Diaz et al. 2009, Smith et al. 2012</p>	<p><b>lung alveola:</b> Huh et al. 2010</p> <p><b>liver lobulus:</b> Kane et al. 2006, Hwa et al. 2007, Khetani et al. 2008</p> <p><b>small artery:</b> Günther et al. 2010</p> <p><b>intestinal villus:</b> Ootani et al. 2010, Sato et al. 2009, Sung et al. 2011, Lahar et al. 2011, Yu et al. 2012</p> <p><b>nervous system:</b> Park et al. 2009</p> <p><b>bone-marrow units:</b> Cui et al. 2007</p>
<b>multiple</b>	<p><b>bone-marrow+liver+tumour:</b> Viravaidya et al 2004, Tabosian et al 2009, Sung et al 2009 and 2010, Mahler et al 2009 and 2012</p> <p><b>lung+liver+kidney+adipose:</b> Zhang et al. 2009</p> <p><b>intestine+liver+tumour:</b> Imura et al. 2010</p>	

*ATLA* 2012, 40, 235-257 Marx et al: ‘Human-on-a-chip’ developments: A translational cutting edge alternative to systemic safety assessment and efficiency evaluation of substances in laboratory animals and man?

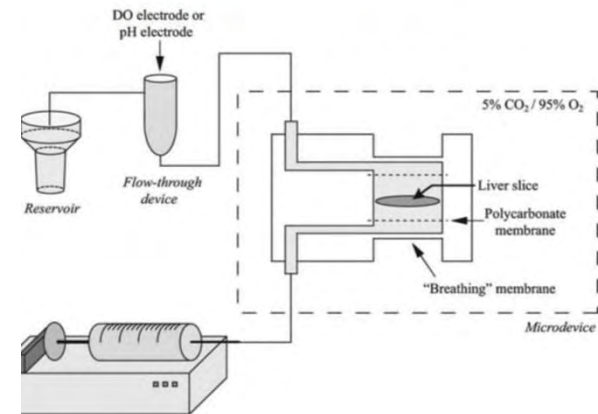
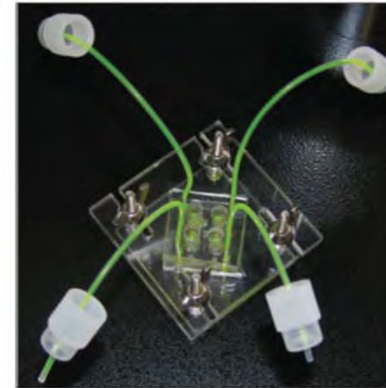
# Perfused liver models

## LiverChip™



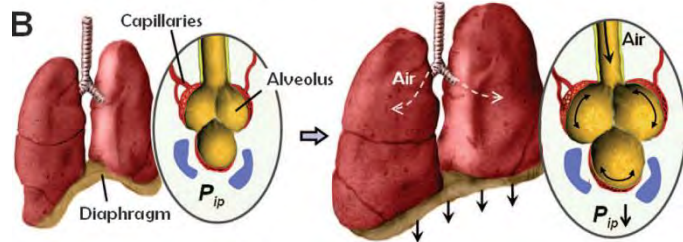
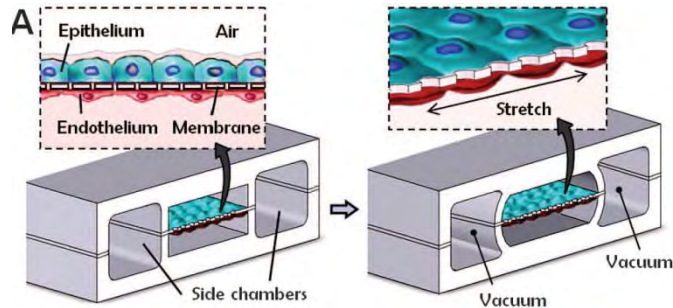
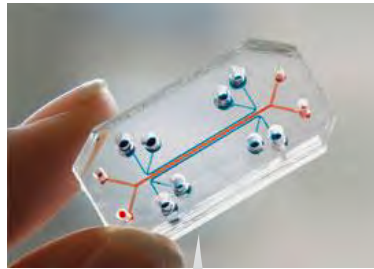
Domansky et al., Lab Chip, 2010, 10, 51-8

## BioChip for liver slices

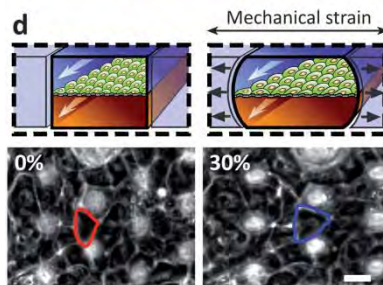
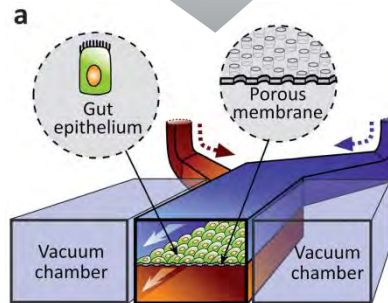


van Midwoud et al., Biotechnol. Bioeng., 2010, 105, 184-94

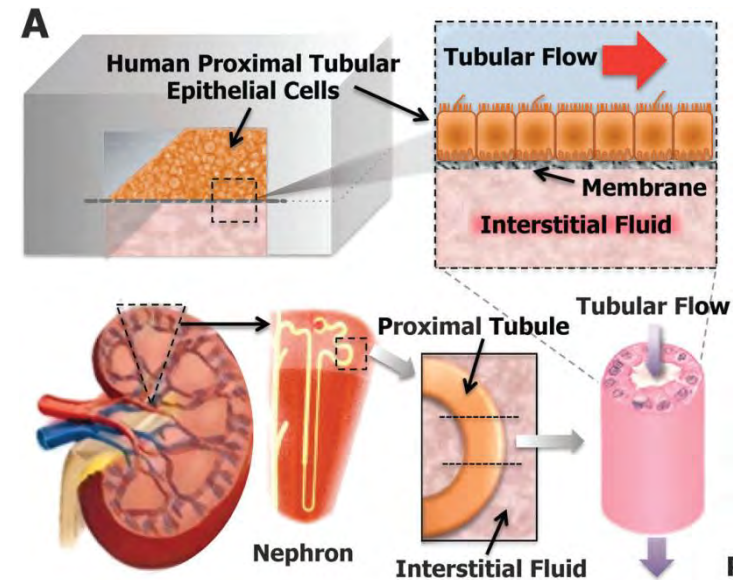
# The Wyss Institute – Organs-on-a-Chip



Huh et al., Science, 2010, 328, 1662



Kim et al., Lab Chip, 2012, 328, 1662



Jang et al., Integr. Biol., 2013, 5, 119

# Artery-on-a-Chip

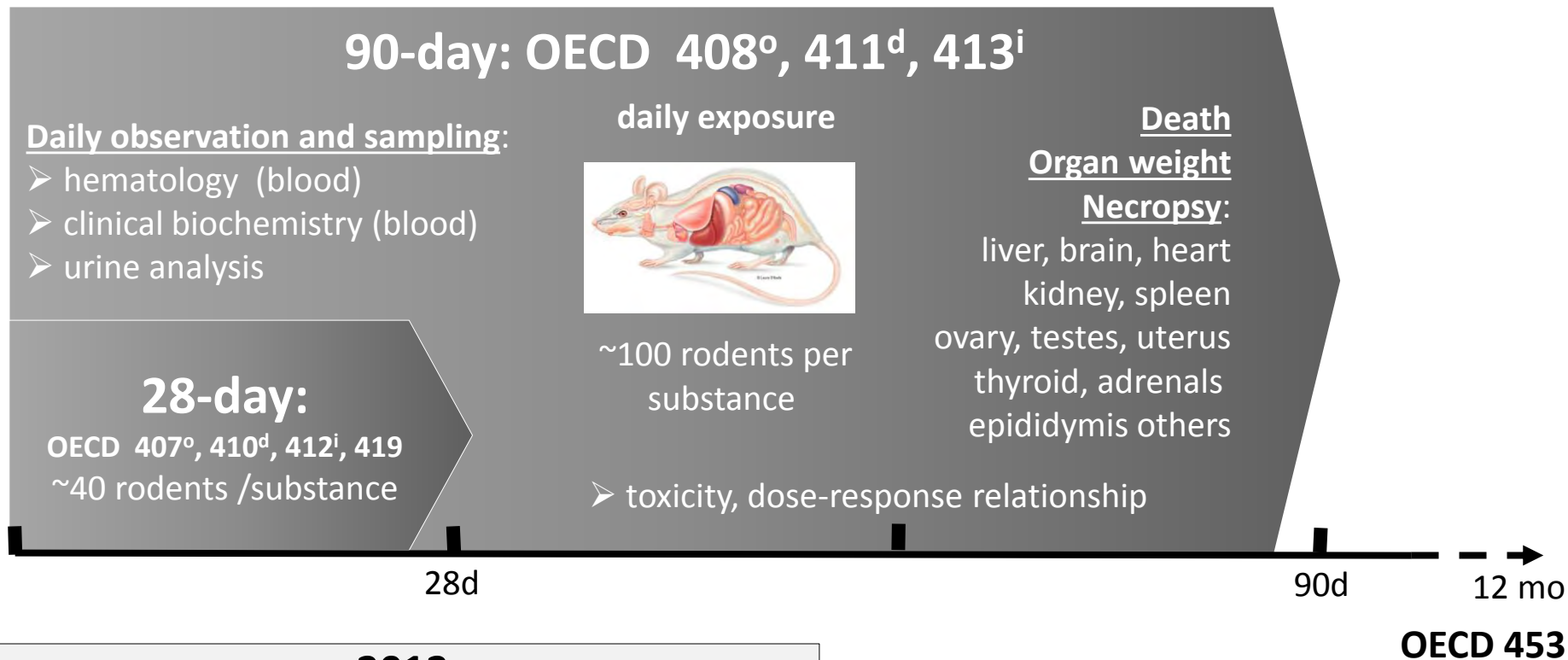
A microfluidic platform for probing small artery structure and function.



Günther et al., Lab Chip, 2010, 10, 2341



# Repeated dose systemic toxicity testing



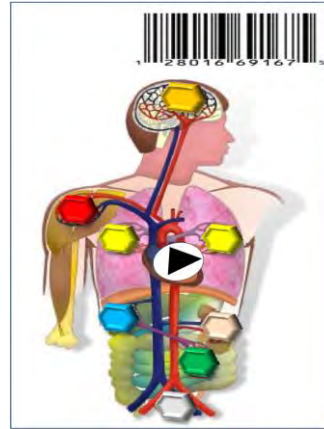
**2012**

**A roadmap for the development of alternative (non-animal) methods for systemic toxicity testing**

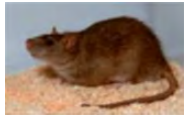
Basketter et al, ALTEX 29, 1/12, pp 1-91

➔ **Integrated Testing Strategy (ITS)**

# Solving the Substance Testing Dilemma



**“Human-on-a-Chip”  
human AND systemic**



animal models

systemic but **NOT** human

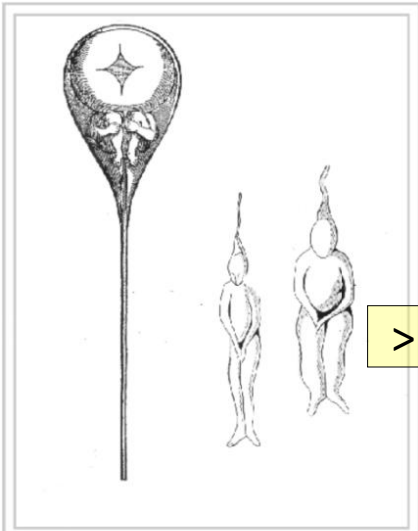


static 2D & 3D

human cell culture

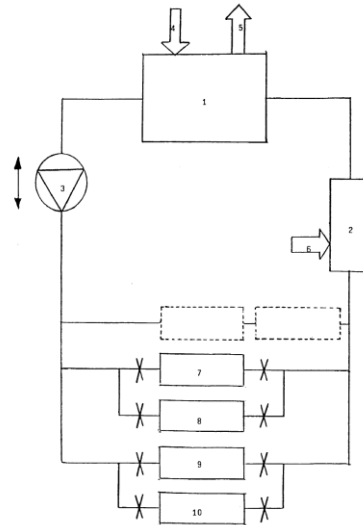
human but **NOT** systemic

# A historic imagination turned into a vision!



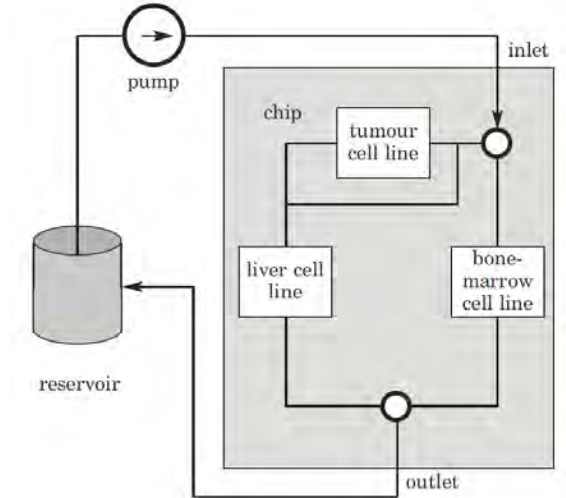
Homunculi in sperm as drawn by N. Hartsoecker in 1695

> 300 Years



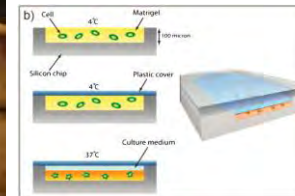
Marx et al, 1992  
V&V zur zeitgleichen Kultivierung  
unterschiedlicher Säugerzellen  
EP0584170B1

- Norecopa -



$\mu$ CCA

Micro cell culture analog



Michael Shuler et al, 2004  
„Human-on-a-chip“  
Biotech. Progress 20, 590-597

# „Human-on-a-chip“ – historical sketch

Michael Shuler et al,  
Cornell University



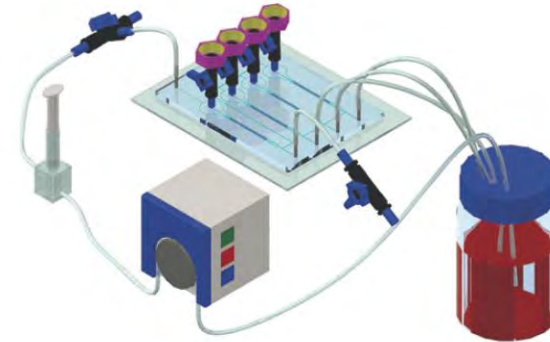
Hanry Yu et al,  
Singapore – US-MIT Alliance



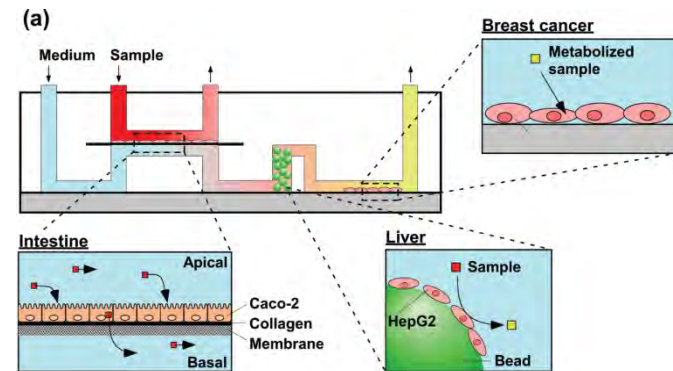
Kiichi Sato et al,  
University of Tokyo



Uwe Marx et al,  
Technische Universität Berlin



Zhang et al., Lab Chip, 2009, 9, 3185

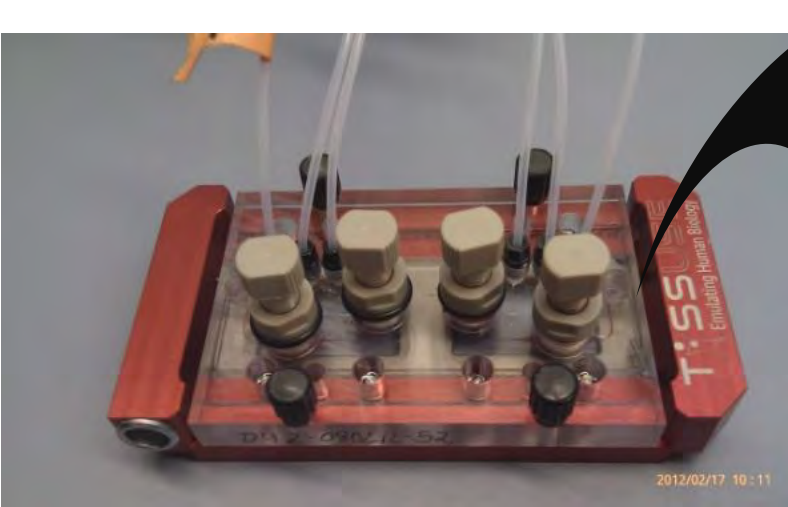


Imura et al., Anal. Chem., 2010, 82, 8

**Nature** March 31<sup>st</sup> 2011: Vol 471 pp. 661-665 M. Baker: Technology feature: A living system on a chip

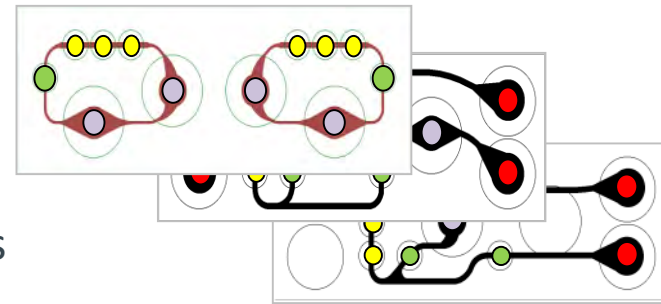
**ATLA** 2012, 40, 235-257 Marx et al: 'Human-on-a-chip' developments: A translational cutting edge alternative to systemic safety assessment and efficiency evaluation of substances in laboratory animals and man?

# The Multi-Organ-Chip (MOC) Technology

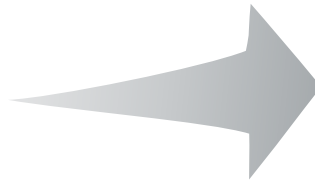
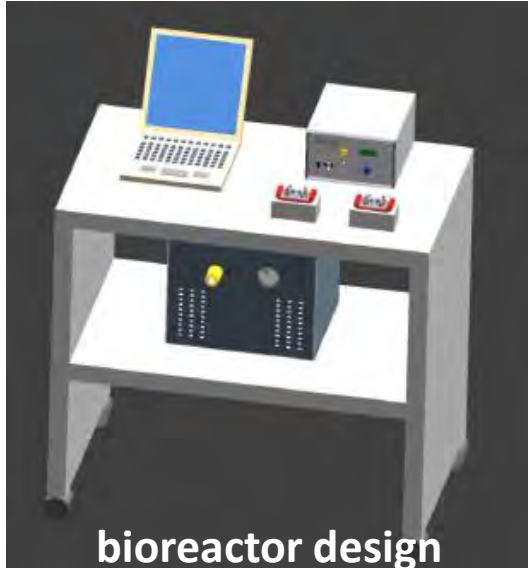


## Features:

- Chip format of a standard microscopic slide
- On-chip micro-pump and natural tissue to fluid ratio
- Variable physiological shear stresses applicable
- Tissue cultures 100,000-fold smaller than original organs
- Rapid prototyping of any relevant chip design
- Compatible with life tissue imaging - Norecopa -

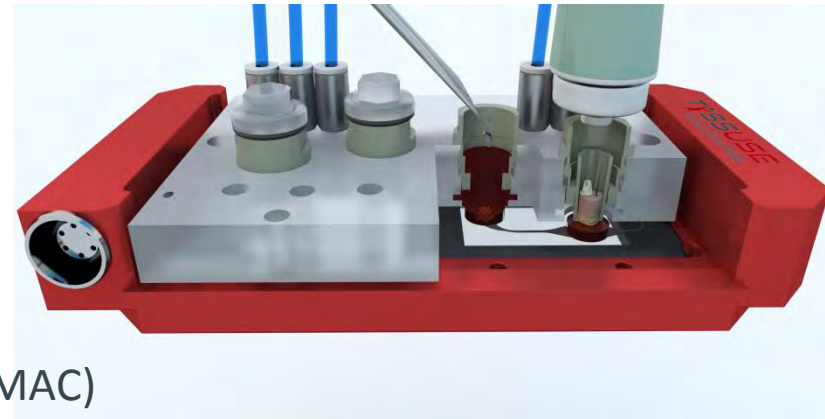


# Benchtop bioreactor



## Features:

- Controlling up to 24 pneumatic actors
- Up to 4 chips per system
- Adjustable temperature and fluid flow
- Software control (e.g. WINDOWS, LINUX, MAC)
- Telemonitoring

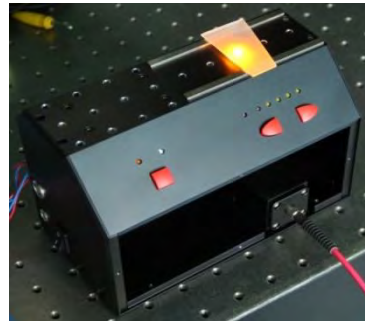


# Sensors / In-process-controls

parameter approach	flow velocity	organ viability	organ functionality	pH & pO <sub>2</sub>	t°
principle	particle imaging velocimetry	fluorescence spectroscopy	surface plasmon resonance for secreted proteins	fluorescence lifetime	PT1000 temperature detector
features	non invasive different spots biological particles	cell tracker live imaging double staining possible	multiple proteins (46 per micro sensor 10 mm x 0.8 mm)	fibre coupled external calibration	long-term robustness

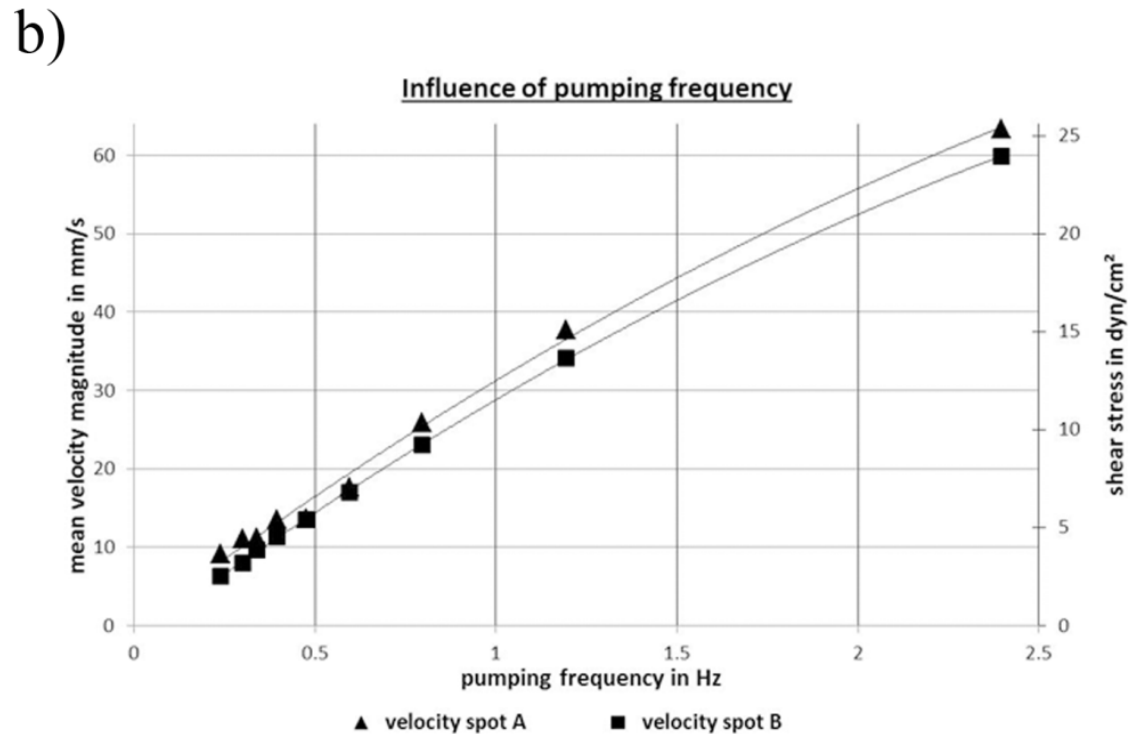
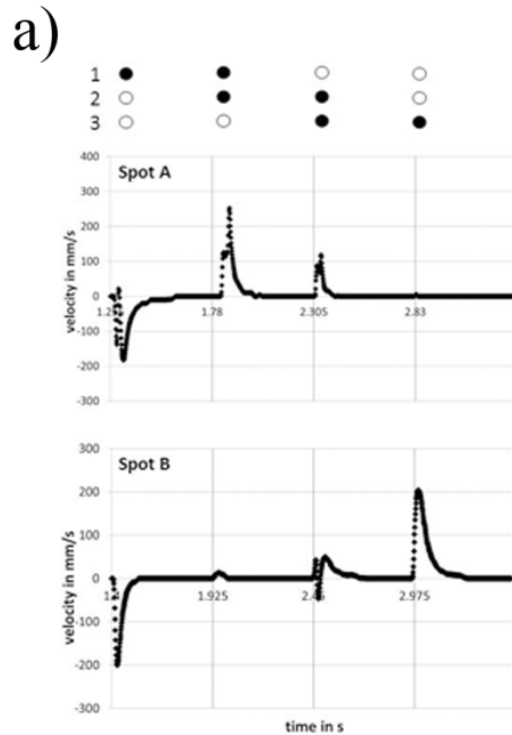


Frank Sonntag



# Evaluation of fluid dynamics

## Particle imaging velocimetry



Schimek et al., Lab Chip 2013



# Smallest possible scale of organs

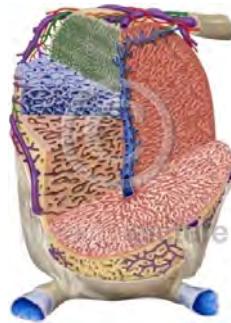
- a single liver lobule is of 1,3  $\mu\text{l}$  in scale
- 1 million liver lobules constitute a human liver



Molecule



Cell



Organoid



Organ

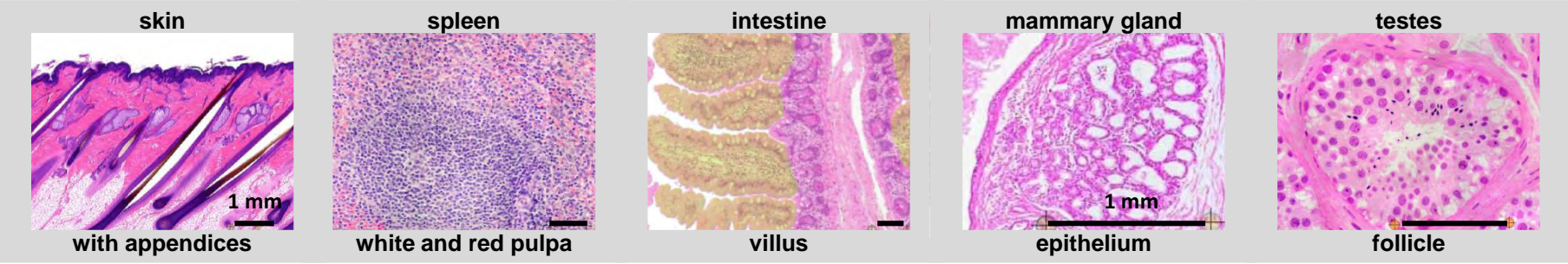


Individual

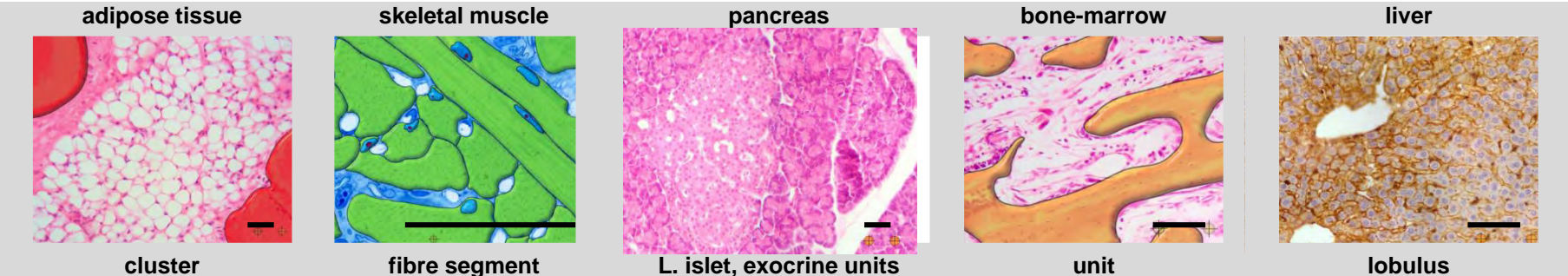
Ten liver lobules – the basis for a  $\frac{1}{100.000}$  “human-on-a-chip”

# Human organoids

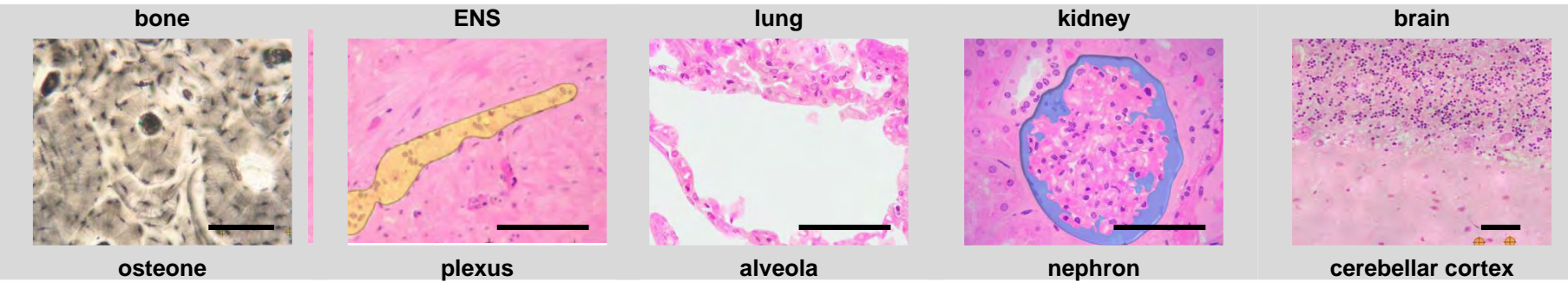
High turnover  
High regenerative potential



Low turnover  
High regenerative potential



Low turnover  
Low regenerative potential

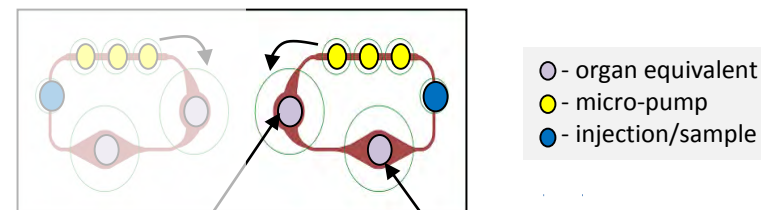
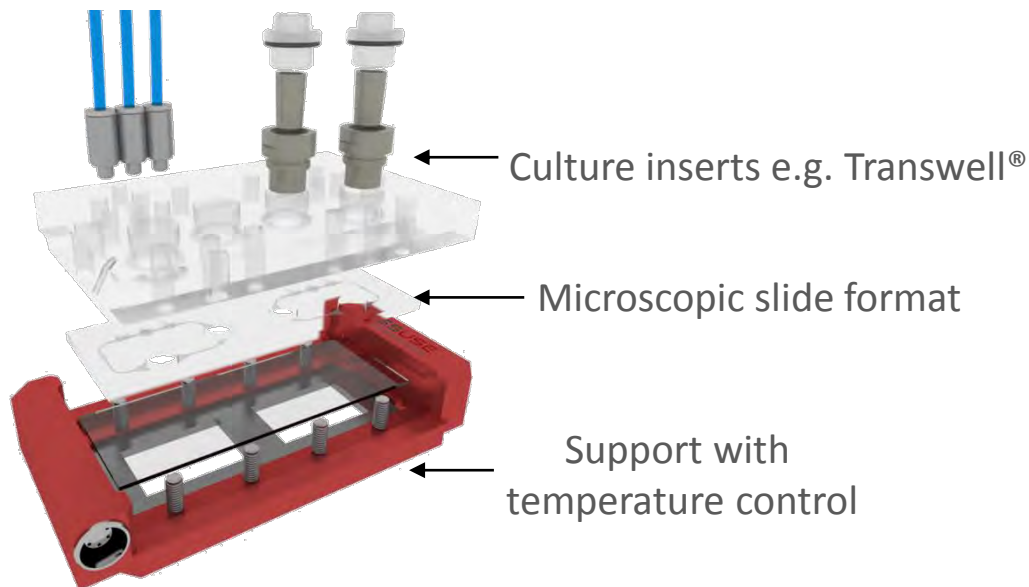


Bars: 100  $\mu$ m

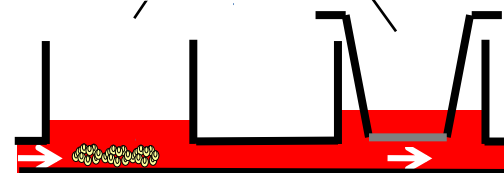
Marx et al., Altern Lab Anim. 2012 Oct;40(5):235-57

- Norecopa -

# The “Two-Tissue Culture Chip”



- - organ equivalent
- - micro-pump
- - injection/sample



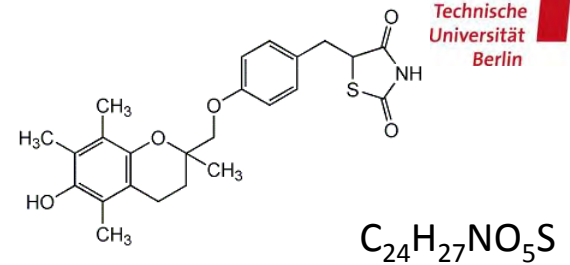
equivalent to  
10 liver lobuli

cardiac, neuronal,  
Immune tissue  
bone marrow  
skin, intestine  
kidney, cancer .....

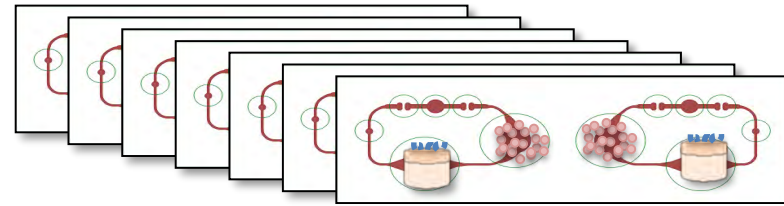
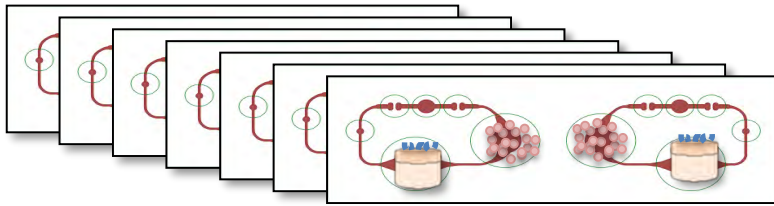
	Duration	Short-term (<48h)	Long-term (<28d)	Homeostasis (90d, 1y...)
Tissue				
liver		✓	✓	in progress
skin		✓	✓	in progress
vasculature		✓	✓	in progress
neurons		✓	✓	in progress
intestine		✓	in progress	in progress
kidney		✓	in progress	in progress

# Tox study using Troglitazone

**Trade name:** Rezulin, Rizulin, Romazin, Sensulin  
**Developed by:** Daiichi Sankyo Co (Japan)  
**Manufactured by:** Parke-Davis (1997 approved by FDA)



**Indication:** Troglitazone is an antidiabetic and anti-inflammatory drug, prescribed for patients with diabetes mellitus type 2  
**Contraindication:** Idiosyncratic reaction leading to liver failure



14 chips comprising **28 circuits** and **20 static controls**.

**Inoculation** of the chips on day 0

Exposition to the drug at varying concentrations. Daily exposure referring to OECD 407.

... **7-14 days experiment**

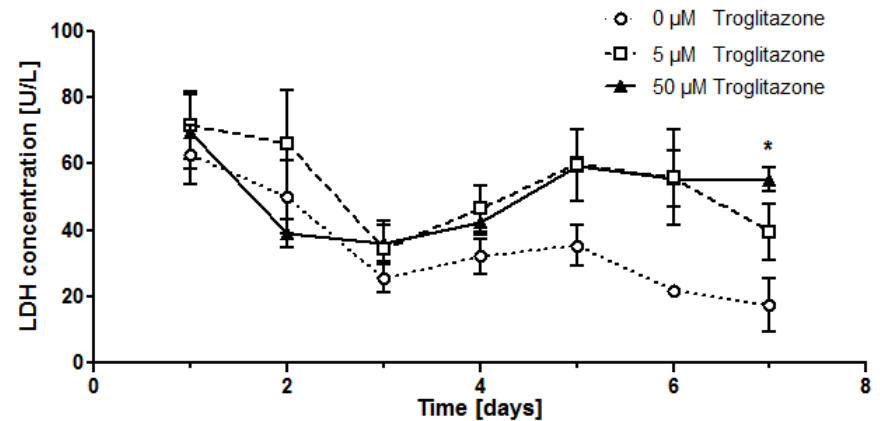
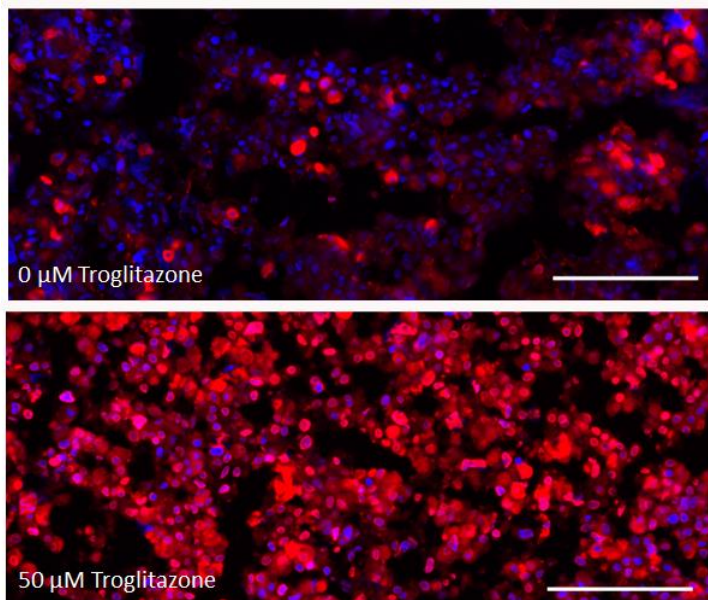
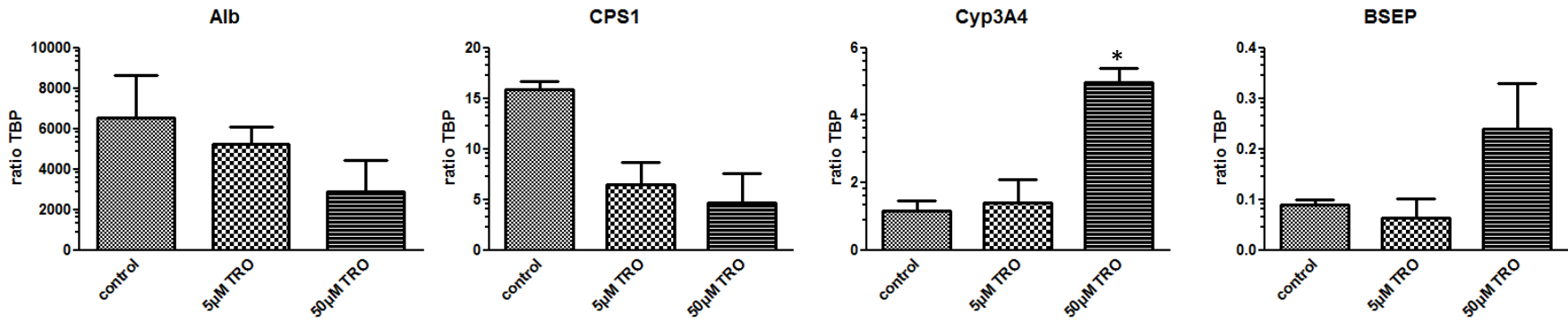
↓ ↓ ↓ ...

Daily media exchange of 250µl.

The supernatants are checked for glucose, lactate, pH, albumin and LDH

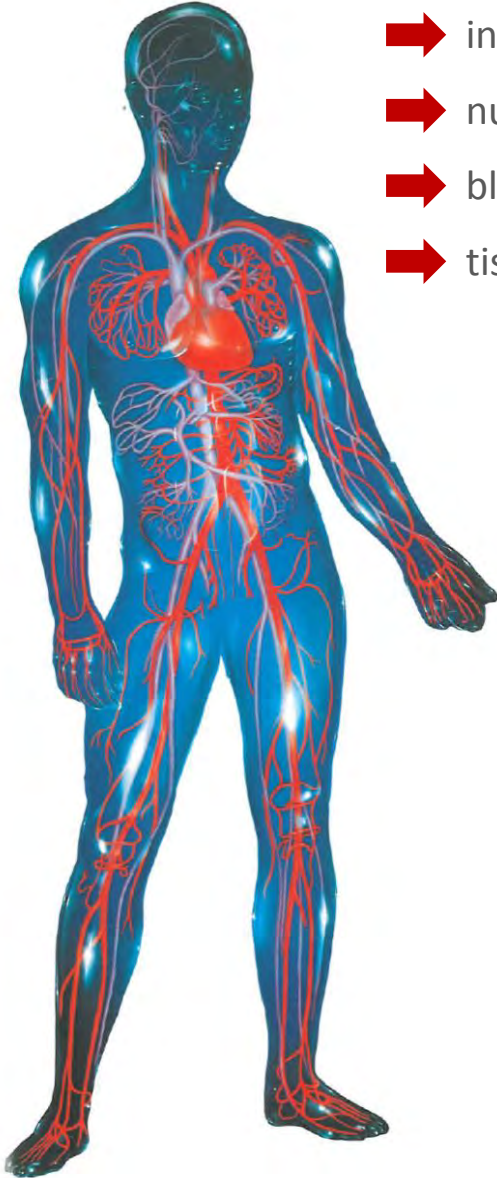
↓  
Endpoint analysis by IHC and RT-PCR

# Sensitivity to Troglitazone (7 day exposure)

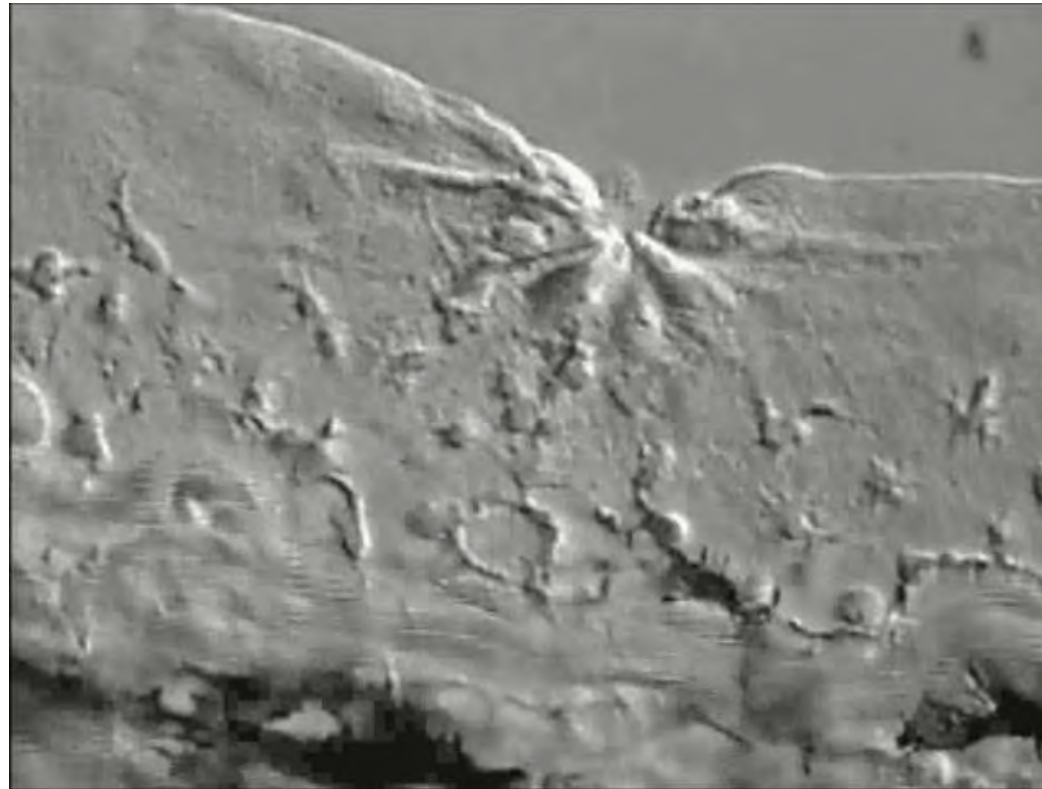


Wagner et al., Lab Chip 2013

# The crucial role of dynamic blood circulation

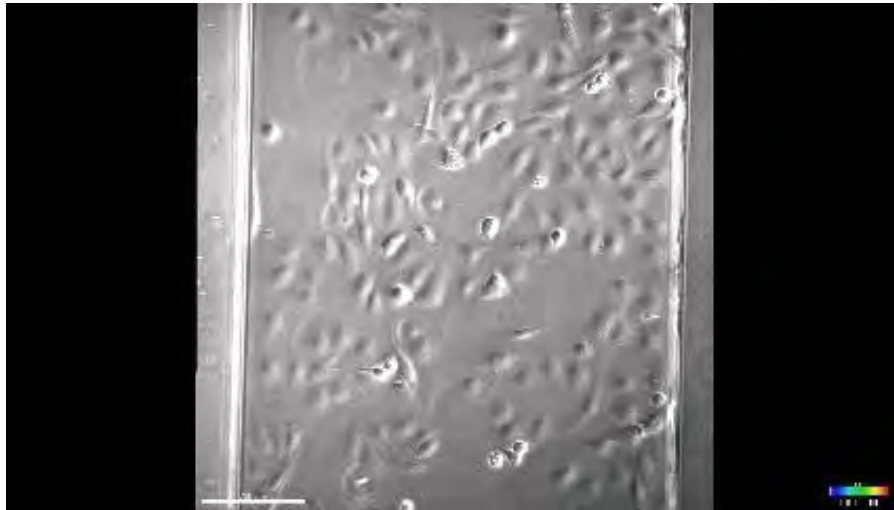


- ➔ interconnection of organs to create an **organism**
- ➔ nutrient and oxygen transport through **blood plasma and red blood cells**
- ➔ blood-tissue barrier and neo angiogenesis through **endothelial cells**
- ➔ tissue repair and immune response through **white blood cells**



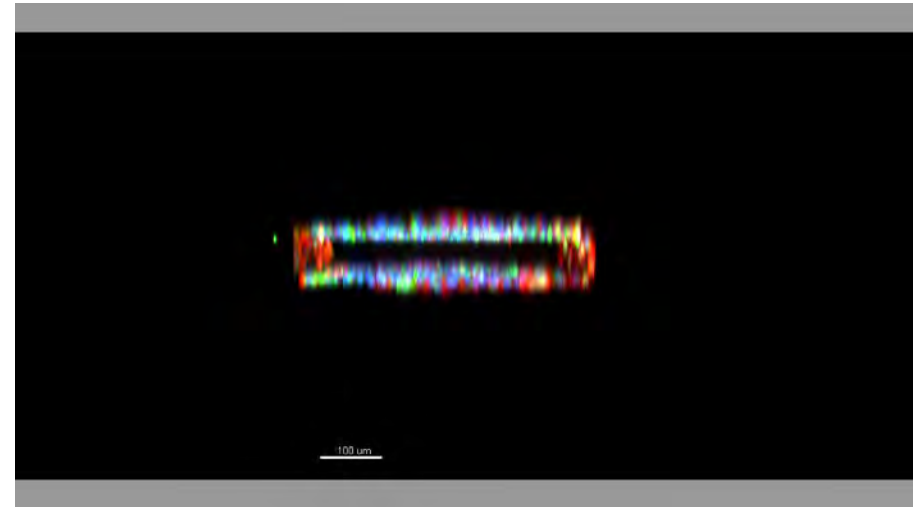
# Establishment of stable microvascular circuits

## Live-cell imaging



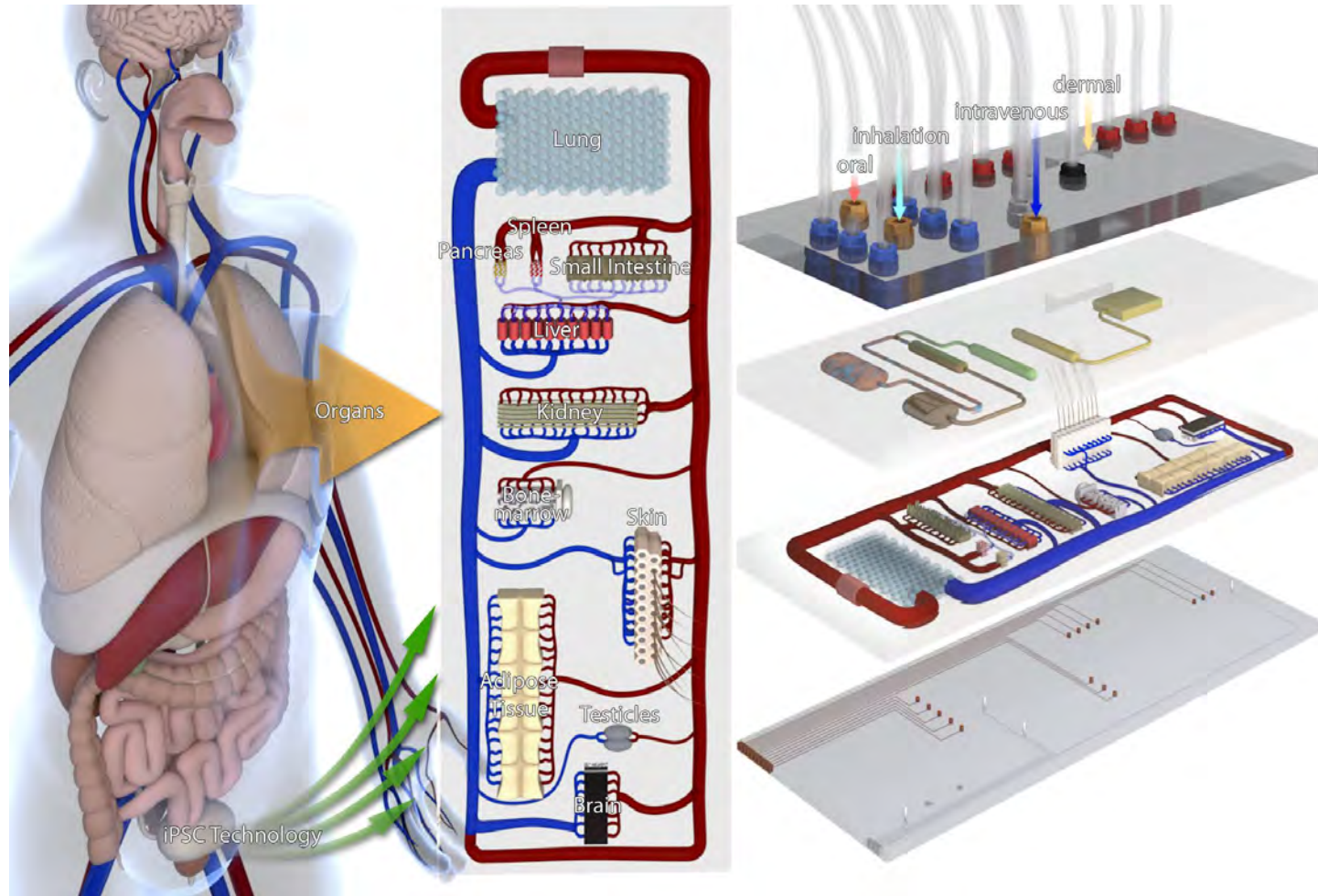
Human microvascular endothelial cells  
(66h - time lapse; scale bar: 200 $\mu$ m)

## Endpoint control



Human microvascular endothelial cells  
cultured for 3 days under constant shear stress  
(von Willebrand-Faktor: green; CD31: red;  
Nuclei: blue; scale bar: 200 $\mu$ m)

# Next generation Multi-Organ-Chip



Marx et al., Altern Lab Anim. 2012 Oct;40(5):235-57

- Norecopa -



# US Initiative “Human-on-chip”

US-Initiative adopted the “human-on-a-chip” strategy 2012 in a unique way

FDA

July 1<sup>st</sup> 2012

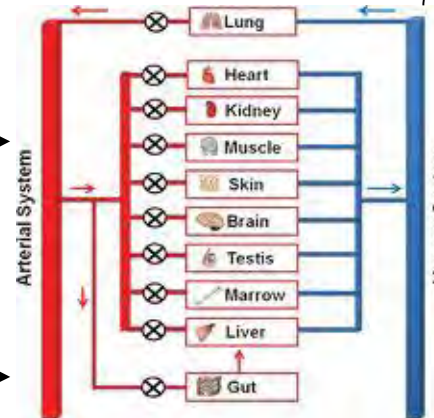
June 30<sup>th</sup> 2017

DARPA  
Defence Advanced  
Research Projects Agency

US\$ 69Mio/5 Years

NIH/NCATS  
National Center for Advancing  
Translational Sciences

US\$ 64Mio/5 Years



17 NIH groups and ...



Linda Griffith  
MIT



Donald Ingber  
Wyss Institute  
(Harvard MS)

## Three players – NIH, FDA, DARPA

Source: The Burrill Report. [http://www.burrillreport.com/article-nih\\_and\\_darpa\\_fund\\_development\\_of\\_organ\\_on\\_a\\_chip\\_systems.html](http://www.burrillreport.com/article-nih_and_darpa_fund_development_of_organ_on_a_chip_systems.html)

Adopted from Suzanne Fitzpatrick, FDA 27.10.2012

# Thank you for your attention!



Ilka Wagner, Eva-Maria Materne, Lutz Kloke, Chris Drewell, Katharina Schimek, Tobias Hasenberg, Silke Hoffmann, Gerd Lindner, Juliane Hübner, Alexandra Lorenz, Caroline Frädrich, Annika Jaenicke, Agnes Schumacher, Luzie Reiners-Schramm, Jennifer Binder, Shirin Fatehi, Mark Rosowski, Beren Atac, Marielle Königsmark, Sandro Wagner, Karolina Tykwinska, Özlem Vural, Manuela Peters, Alexander Thomas, Roland Lauster, Uwe Marx